

101.428-12 – including *Taq* polymerase, IFU-01
101.428-12u – without *Taq* polymerase, IFU-02

Visit <https://labproducts.caredx.com> for
“Instructions for Use” (IFU)

Lot No.: **5L0**

Lot-specific Information
Olerup SSP[®] HLA-A*29

Product number:	101.428-12 – including <i>Taq</i> polymerase 101.428-12u – without <i>Taq</i> polymerase
Lot number:	5L0
Expiry date:	2024-08-01
Number of tests:	12
Number of wells per test:	30+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 5L0.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP[®]
HLA-A*29 LOT (3H6)**

- The product documentation has been updated for new alleles of IMGT 3.40.0.
- The kit resolution focuses on common and well documented (CWD) alleles¹.
- One new well has been added to HLA-A*29, well **31**.

¹As described in section Uniquely Identified Alleles.

The HLA-A*29 primer set, specificity and interpretation tables have been updated for the HLA-A alleles described since the previous *Olerup SSP[®] HLA-A*29* lot was made (**Lot No. 3H6**).

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
30	Added, moved	Added, moved	Primer pair added for the A*29:88 allele. Negative control moved to primer mix 31.
31	Added	Added	Negative control added from primer mix 30.

¹S. J. Mack, P. Cano, J. A. Hollenbach et al.
Common and well-documented HLA alleles: 2012 update to
the CWD catalogue. Tissue Antigens, 2013, 81, 194–203

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Well **31** contains Negative Control primer pairs, that will amplify the majority of the *Olerup* SSP[®] HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.
 The PCR product generated by the positive control primer pair is 200 base pairs.

Length of PCR product	105	200	105	80	75	80	85
5'-primer¹	164	340	440	45	45	43	36
	5'-CAC ^{3'}	5'-Agg ^{3'}	5'-TTA ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-TAC ^{3'}
							36
							5'-TAT ^{3'}
3'-primer²	231	2nd I	507	59	58	57	47
	5'-TgC ^{3'}	5'-AAA ^{3'}	5'-TTg ^{3'}	5'-CTC ^{3'}	5'-ggC ^{3'}	5'-CTC ^{3'}	5'-ACA ^{3'}
							48
							5'-gCA ^{3'}
							48
							5'-gCC ^{3'}
							52
							5'-TgT ^{3'}
A*	+	+	+				
B*	+	+	+				
C*	+	+	+				
DRB1				+	+		
DRB3				+	+		
DRB5				+			
DQB1					+		
DPB1						+	
DQA1							+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot-specific Information
PRODUCT DESCRIPTION

HLA-A*29 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the A*29:01 to A*29:144 alleles.

PLATE LAYOUT

Each test consists of 31 PCR reactions in a 32 well cut PCR plate. Well 32 is empty.

1	2	3	4	5	6	7	8
9	10	11	12	13	14	15	16
17	18	19	20	21	22	23	24
25	26	27	28	29	30	NC	empty

The 32 well cut PCR plate is marked with ‘HLA-A*29’ in silver/gray ink.

Well No. 1 is marked with the Lot No. ‘5L0’.

Wells 1 to 30 – HLA-A*29 high resolution primers.

Well 31 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded. The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 32 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between HLA-A alleles, non-HLA-A*29 alleles will be amplified by some primer mixes. For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the HLA-A*29 alleles, i.e. **A*29:01 to A*29:144 alleles**, recognized by the HLA Nomenclature Committee in April 2020^{1,2} will be amplified by the primers in the HLA-A*29 subtyping kit.

The HLA-A*29 kit enables separation of the confirmed HLA-A*29 alleles as listed in the IMGT/HLA database 3.29.0. An HLA allele is listed as confirmed by IMGT/HLA if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for HLA-A*29 alleles is listed below.

The HLA-A*29 kit also enables identification of many null and alternatively expressed alleles.

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Lot-specific Information

The following HLA-A*29 alleles can be distinguished by the different sizes of the HLA-specific PCR product:

Alleles	Primer mix
A*29:07, 29:46	8
A*29:08N, 29:139	9
A*29:11, 29:92	13
A*29:17, 29:144	10
A*29:20, 29:82	29

¹HLA-A alleles listed on the IMGT/HLA web page 2020-April-20, release 3.40.0, www.ebi.ac.uk/imgt/hla.

²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

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Lot-specific Information

ALLELE CONFIRMATION STATUS

Allele	Status ¹	Allele	Status ¹	Allele	Status ¹	Allele	Status ¹
A*29:01:01:01	Confirmed	A*29:08N	Unconfirmed	A*29:47	Unconfirmed	A*29:87	Confirmed
A*29:01:01:02N	Unconfirmed	A*29:09	Confirmed	A*29:48	Unconfirmed	A*29:88	Unconfirmed
A*29:01:02	Unconfirmed	A*29:10:01	Confirmed	A*29:49	Unconfirmed	A*29:89	Unconfirmed
A*29:01:03	Unconfirmed	A*29:10:02	Confirmed	A*29:50	Unconfirmed	A*29:90	Confirmed
A*29:01:04	Unconfirmed	A*29:11	Confirmed	A*29:51	Confirmed	A*29:91	Unconfirmed
A*29:01:05	Unconfirmed	A*29:12	Confirmed	A*29:52	Unconfirmed	A*29:92	Unconfirmed
A*29:01:06	Unconfirmed	A*29:13	Unconfirmed	A*29:53	Confirmed	A*29:93	Unconfirmed
A*29:01:07	Unconfirmed	A*29:14	Unconfirmed	A*29:54	Confirmed	A*29:94	Unconfirmed
A*29:01:08	Unconfirmed	A*29:15	Unconfirmed	A*29:55	Confirmed	A*29:95	Unconfirmed
A*29:01:09	Unconfirmed	A*29:16	Unconfirmed	A*29:56	Unconfirmed	A*29:96	Unconfirmed
A*29:02:01:01	Confirmed	A*29:17	Unconfirmed	A*29:57	Unconfirmed	A*29:97	Unconfirmed
A*29:02:01:02	Confirmed	A*29:18	Unconfirmed	A*29:58	Unconfirmed	A*29:98	Unconfirmed
A*29:02:01:03	Confirmed	A*29:19	Unconfirmed	A*29:59	Unconfirmed	A*29:99	Unconfirmed
A*29:02:01:04	Confirmed	A*29:20	Confirmed	A*29:60	Unconfirmed	A*29:100	Unconfirmed
A*29:02:02	Confirmed	A*29:21	Confirmed	A*29:61	Unconfirmed	A*29:101	Unconfirmed
A*29:02:03	Confirmed	A*29:22	Confirmed	A*29:62	Unconfirmed	A*29:102	Unconfirmed
A*29:02:04	Confirmed	A*29:23	Unconfirmed	A*29:63	Unconfirmed	A*29:103	Confirmed
A*29:02:05	Unconfirmed	A*29:24	Unconfirmed	A*29:64	Unconfirmed		
A*29:02:06	Confirmed	A*29:25	Unconfirmed	A*29:65	Unconfirmed		
A*29:02:07	Unconfirmed	A*29:26	Confirmed	A*29:66	Unconfirmed		
A*29:02:08	Unconfirmed	A*29:27	Unconfirmed	A*29:67	Unconfirmed		
A*29:02:09	Confirmed	A*29:28	Unconfirmed	A*29:68	Unconfirmed		
A*29:02:10	Unconfirmed	A*29:29	Unconfirmed	A*29:69	Confirmed		
A*29:02:11	Unconfirmed	A*29:30	Unconfirmed	A*29:70	Unconfirmed		
A*29:02:12	Unconfirmed	A*29:31	Unconfirmed	A*29:71	Confirmed		
A*29:02:13	Unconfirmed	A*29:32	Confirmed	A*29:72	Unconfirmed		
A*29:02:14	Confirmed	A*29:33	Unconfirmed	A*29:73	Unconfirmed		
A*29:02:15	Unconfirmed	A*29:34	Confirmed	A*29:74	Unconfirmed		
A*29:02:16	Confirmed	A*29:35	Confirmed	A*29:75	Unconfirmed		
A*29:02:17:01	Unconfirmed	A*29:36	Confirmed	A*29:76	Unconfirmed		
A*29:02:17:02	Unconfirmed	A*29:37	Unconfirmed	A*29:77	Confirmed		
A*29:02:18	Unconfirmed	A*29:38	Unconfirmed	A*29:78N	Unconfirmed		
A*29:02:19	Unconfirmed	A*29:39	Unconfirmed	A*29:79	Unconfirmed		
A*29:02:20	Confirmed	A*29:40	Confirmed	A*29:80	Unconfirmed		
A*29:02:21	Unconfirmed	A*29:41	Unconfirmed	A*29:81	Confirmed		
A*29:03	Confirmed	A*29:42	Unconfirmed	A*29:82	Confirmed		
A*29:04	Unconfirmed	A*29:43	Confirmed	A*29:83	Unconfirmed		
A*29:05	Unconfirmed	A*29:44	Confirmed	A*29:84	Unconfirmed		
A*29:06	Unconfirmed	A*29:45	Unconfirmed	A*29:85	Unconfirmed		
A*29:07	Unconfirmed	A*29:46	Unconfirmed	A*29:86	Unconfirmed		

¹Allele status “confirmed” or “unconfirmed” as listed on the IMGT/HLA web page 2017-August-10, release 3.29.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in HLA-A*29 homo- and heterozygotes is available upon request.

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Lot-specific Information
SPECIFICITY TABLE

HLA-A*29 SSP subtyping

Specificities and sizes of the PCR products of the 30+1 primer mixes used for HLA-A*29 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified HLA-A*29 alleles ³	Other amplified HLA-A alleles
1	480 bp	800 bp	*29:01:01:01-29:01:06, 29:01:08-29:01:12, 29:12, 29:15-29:17, 29:20, 29:24, 29:28, 29:33, 29:48-29:49, 29:55-29:58, 29:60-29:62, 29:67, 29:71, 29:76-29:77, 29:79, 29:81-29:83, 29:98-29:99, 29:101:01-29:101:02, 29:107, 29:110, 29:113, 29:115, 29:117-29:118, 29:123-29:125, 29:144	
2	130 bp	1070 bp	*29:01:01:02N	
3	440 bp	800 bp	*29:02:01:01-29:02:03, 29:02:05-29:11, 29:13-29:14, 29:19, 29:21-29:23, 29:26-29:27, 29:29-29:32, 29:34-29:47, 29:50-29:54, 29:59, 29:63, 29:65-29:66, 29:68-29:70, 29:72-29:75, 29:78N, 29:80, 29:84-29:97, 29:100, 29:102-29:106, 29:108, 29:111-29:112N, 29:114, 29:116, 29:119-29:122, 29:126Q-29:135, 29:137-29:143	*01:69:03, 11:01:42, 30:01:11, 30:152, 68:130:02
4⁴	110 bp 165 bp	800 bp	*29:35, 29:51, 29:69, 29:73 *29:03, 29:33	*02:24:02, 02:507, 32:109 *01:01:114, 23:03:01, 24:21:03, 24:208:01, 31:05, 32:13, 33:10
5	130 bp 185 bp	1070 bp	*29:04 *29:23	
6	130 bp	1070 bp	*29:05, 29:33, 29:40, 29:77, 29:87, 29:104	*11:01:28, 11:01:77, 24:21:03, 24:208:01, 31:24, 31:136, 32:02, 32:22, 33:59, 33:102, 33:150, 33:190
7	210 bp	1070 bp	*29:06, 29:123 ^w	*31:51 ^w , 32:12 ^w , 33:168 ^w , 74:26 ^w , B*08:01:07^w , B*15:02:07^w , B*15:17:03^w , B*27:07:05^w , B*41:01:05^w , B*55:02:10^w , C*02:02:15^w , C*03:03:20^w , C*04:175^w , C*07:04:13^w , C*08:01:15^w
8⁴	85 bp	800 bp	*29:07, 29:49	*01:301Q, 03:347, 11:139, 23:53, 23:70, 24:17, 24:41,

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	160 bp		*29:46	24:208:01-24:208:02, 24:488 *32:139
9^{4,5}	80 bp 170 bp	1070 bp	*29:16, 29:139 *29:08N	*01:157, 03:27, 11:233
10^{4,6}	90 bp	800 bp	*29:09, 29:33, 29:51, 29:73, 29:144	*01:01:114, 02:24:02, 02:507, 03:01:18, 11:01:28, 11:01:77, 24:21:03, 24:208:01, 31:24, 32:33:01, 32:109, 33:34, 33:164
	165 bp 215 bp		*29:17, 29:43 *29:54	
11⁴	110 bp 190 bp	800 bp	*29:14, 29:35 *29:10:01-29:10:02, 29:23	C*07:04:13, C*08:01:15
12^{4,5}	80 bp	800 bp	*29:01:11, 29:02:04, 29:18, 29:48	*02:01:165, 03:01:39, 32:01:01:01-32:01:07, 32:01:09-32:01:17, 32:01:19-32:01:29, 32:01:31-32:03:01:02, 32:05-32:55:02, 32:57- 32:69, 32:71, 32:73-32:107, 32:109-32:112N, 32:114- 32:140, 32:142, 68:01:28, 68:01:36, 74:01:01:01- 74:01:05, 74:01:07-74:13, 74:15-74:37
	145 bp		*29:78N	
13^{4,5,7}	90 bp 165 bp 260 bp	1070 bp	*29:11, 29:51, 29:73 *29:12, 29:92 *29:55	*02:24:02, 02:507, 32:109 *31:16, 33:58
14	200 bp	1070 bp	*29:01:01:01-29:13, 29:15- 29:17, 29:19-29:36, 29:38- 29:47, 29:49-29:55, 29:57- 29:104, 29:106-29:144	
15	240 bp	1070 bp	*29:01:01:01-29:12, 29:14- 29:18, 29:20-29:36, 29:38, 29:40-29:47, 29:49-29:55, 29:57-29:97, 29:99-29:144	*01:143, 31:79, 33:13, 33:48
16^{4,5}	95 bp	1070 bp	*29:15	*02:221, 23:41, 30:162, 31:78
	160 bp 190 bp		*29:21, 29:43 *29:53	
17⁴	100 bp 130 bp 190 bp 215 bp	1070 bp	*29:51, 29:69, 29:73 *29:24, 29:40 *29:27, 29:53 *29:54	*02:24:02, 02:507, 32:109
18⁵	225 bp 260 bp	1070 bp	*29:37, 29:56 *29:36	*32:07, 33:119
19⁶	160 bp 260 bp 505 bp	1070 bp	*29:25 *29:55 *29:26	
20^{4,5}	105 bp	1070 bp	*29:44, 29:64	*01:01:114, 02:65, 02:829, 11:01:28, 11:01:77, 31:123, 31:179,

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				32:01:01:01-32:01:06, 32:01:08-32:01:11, 32:01:13-32:01:30, 32:01:32-32:03:01:02, 32:05-32:27N, 32:29- 32:30:01, 32:31-32:33:01, 32:34-32:65, 32:67-32:93, 32:95-32:100, 32:102- 32:128, 32:130N-32:131, 32:133N-32:142, 33:165, 74:01:01:01-74:38
21	185 bp	1070 bp	*29:22	*01:20, 01:66, 01:130, 02:19, 02:44, 02:149, 02:309, 02:408, 02:436, 02:619:01-02:619:02, 02:838, 02:842, 02:927, 03:95, 03:343, 11:288, 24:14:01:01-24:14:01:04, 24:93, 24:324, 26:22, 30:47, 31:99, 33:22, 66:09, B*08:204, C*02:74
22⁴	115 bp	800 bp	*29:29	*01:148, 03:327, 11:128, 26:85, 33:139, 66:41, 68:58:01-68:58:02
	260 bp		*29:13	*24:82, 31:07-31:08, 31:10, 32:42
23⁴	75 bp	1070 bp	*29:32	*02:24:02, 02:65, 02:152, 02:507, 02:829, 23:03:01, 23:83, 24:21:03, 31:01:02:01-31:01:29, 31:01:31-31:02:02, 31:05, 31:07-31:61, 31:63-31:66, 31:70-31:119, 31:121- 31:178, 31:180, 32:01:01:01-32:01:06, 32:01:08-32:01:11, 32:01:13-32:01:27, 32:01:29-32:03:01:02, 32:05-32:27N, 32:29, 32:31, 32:33:01, 32:34- 32:47, 32:49-32:65, 32:67- 32:93, 32:95-32:100, 32:102-32:118, 32:120- 32:142, 33:01:01:01- 33:01:04, 33:01:06- 33:01:14, 33:03:01:01- 33:03:18, 33:03:20- 33:03:23, 33:03:25- 33:03:26, 33:03:28-33:17, 33:20-33:31, 33:33-33:37, 33:39-33:151, 33:153- 33:158, 33:160-33:201, 74:01:01:01-74:38, B*15:17:03
	510 bp		*29:81, 29:133:01:01- 29:133:01:02	

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24⁴	125 bp	1070 bp	*29:71	*31:77
25⁴	110 bp	1070 bp	*29:90	*31:108, 32:120, 33:110
26	185 bp	1070 bp	*29:103	
27	510 bp	1070 bp	*29:81, 29:133:01:01- 29:133:01:02	
28⁴	95 bp	1070 bp	*29:112N	
29⁴	105 bp 475 bp	1070 bp	*29:19-29:20, 29:34 *29:82	*01:69:03
30	270 bp	1070 bp	*29:88	
31⁸	-	-	Negative Control	

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of HLA-A*29 SSP typings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several HLA Class I alleles 1st and/or 4th exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

⁵Primer mixes 9, 12, 13, 16, 18 and 20 may have tendencies of unspecific amplifications.

⁶Primer mixes 10 and 19 have a tendency giving rise to primer oligomer formation.

⁷Primer mix 13 may give rise to a lower yield of HLA-specific PCR product than the other A*29 primer mixes.

⁸Primer mix 31 contains a negative control, which will amplify the majority of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 200 base pairs.

Abbreviations

w: might be weakly amplified.

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PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	480	130	440	110	130	130	210	85	80	90	110	80
				165	185			160	170	165	190	145
										215		
Length of int. pos. control ¹	800	1070	800	800	1070	1070	1070	800	1070	800	800	800
5'-primer(s) ²	180	808	219	448	180	448	448	368	97	82	180	180
	5'-TTT 3'	5'-CgT 3'	5'-gCA 3'	5'-CCT 3'	5'-TTT 3'	5'-CCT 3'	5'-CCT 3'	5'-gTT 3'	5'-TCA 3'	5'-ACC 3'	5'-TTT 3'	5'-TTT 3'
								652	413	130	448	
								5'-CTg 3'	5'-CCg 3'	5'-AgA 3'	5'-CCT 3'	
										140		
										5'-CAA 3'		
										448		
										5'-CCT 3'		
3'-primer(s) ³	376	895	376	506	268	533	616	413	224	257	238	218
	5'-gTg 3'	5'-CTC 3'	5'-gTC 3'	5'-TgT 3'	5'-ATg 3'	5'-gCC 3'	5'-CgC 3'	5'-gCC 3'	5'-TCT 3'	5'-gCA 3'	5'-CCT 3'	5'-gCg 3'
				526	326	539		773	454	497	326	286
				5'-CAT 3'	5'-TgA 3'	5'-TCT 3'		5'-gCT 3'	5'-CTg 3'	5'-Tgg 3'	5'-TgA 3'	5'-CTA 3'
				570						502	526	
				5'-CCg 3'						5'-CTT 3'	5'-CAT 3'	
											601	
											5'-CTT 3'	
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15	16	17	18	19	20	21	22	23	24
Length of spec. PCR product	90	200	240	95	100	225	160	105	185	115	75	125
	165			160	130	260	260			260	510	
				190	190		505					
					215							
Length of int. pos. control ¹	1070	1070	1070	1070	1070	1070	1070	1070	1070	800	1070	1070
5'-primer(s) ²	97	98	98	107	82	41	98	385	355	98	221	448
	5'-TCA 3'	5'-CAC 3'	5'-CAC 3'	5'-CgC 3'	5'-ACC 3'	5'-CTT 3'	5'-CAC 3'	5'-ggC 3'	5'-CCg 3'	5'-CAC 3'	5'-ACA 3'	5'-CCT 3'
	448			134	107		3 rd I			355	413	
	5'-CCT 3'			5'-CCT 3'	5'-CgC 3'		5'-ATA 3'			5'-CCA 3'	5'-CCA 3'	
				140	448							
				5'-CAA 3'	5'-CCT 3'							
				484								
				5'-ACg 3'								
3'-primer(s) ³	221	257	299	257	257	97	217	448	497	317	448	530
	5'-ACA 3'	5'-gCA 3'	5'-TCg 3'	5'-gCA 3'	5'-gCA 3'	5'-ggA 3'	5'-TgA 3'	5'-CAA 3'	5'-TgA 3'	5'-ggA 3'	5'-CAA 3'	5'-CCA 3'
	316			538	506	131	316			430		
	5'-gCT 3'			5'-CAA 3'	5'-TgT 3'	5'-ggA 3'	5'-gCT 3'			5'-gCT 3'		
	494			533			667					
	5'-TCg 3'			5'-gCC 3'			5'-ggT 3'					
	497			545								
	5'-Tgg 3'			5'-AgC 3'								
				595								
				5'-CCA 3'								
Well No.	13	14	15	16	17	18	19	20	21	22	23	24

101.428-12 – including *Taq* polymerase, IFU-01
 101.428-12u – without *Taq* polymerase, IFU-02

Visit <https://labproducts.caredx.com> for
 “Instructions for Use” (IFU)

Lot No.: **5L0**

Lot-specific Information

Well No.	25	26	27	28	29	30
Length of spec. PCR product	110	185	510	95	105	270
					475	
Length of int. pos. control ¹	1070	1070	1070	1070	1070	1070
5'-primer(s) ²	448	98	221	288	219	98
	5'-CCT 3'	5'-CAC 3'	5'-ACA 3'	5'-..g 3'	5'-gCA 3'	5'-CAC 3'
3'-primer(s) ³	518	242	448	341	282	329
	5'-CCA 3'	5'-CCA 3'	5'-CAA 3'	5'-Cgg 3'	5'-gAg 3'	5'-ggg 3'
					282	
					5'-gAg 3'	
					412	
					5'-CCC 3'	
Well No.	25	26	27	28	29	30

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

101.428-12 – including *Taq* polymerase, IFU-01
 101.428-12u – without *Taq* polymerase, IFU-02

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 “Instructions for Use” (IFU)

Lot No.: **5L0**

Lot-specific Information

CELL LINE VALIDATION SHEET																				
HLA-A*29 SSP subtyping kit ²																				
				Well																
				1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	
				Lot No.:	201901601	201901602	201901603	201901604	201901605	201901606	201901607	201901608	201901609	201901610	201901611	201901612	201901613	201901614	201901615	201901616
	IHWC cell line ¹	A*	A*																	
1	9001 SA	*24:02			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280 LK707	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	9011 E4181324	*01:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275 GU373	*30:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
5	9009 KAS011	*01:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353 SM	*02:01	*26:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020 QBL	*26:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
8	9007 DEM	*31:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
9	9026 YAR	*26:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107 LKT3	*24:02			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
11	9051 PITOUT	*29:02			-	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-
12	9052 DBB	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004 JESTHOM	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071 OLGA	*31:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
15	9075 DKB	*24:02			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037 SWEIG007	*29:02			-	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-
17	9282 CTM3953540	*03:01	*80:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
18	9257 32367	*33:03	*74:01		-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
19	9038 BM16	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
20	9059 SLE005	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
21	9064 AMALA	*02:17			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
22	9056 KOSE	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
23	9124 IHL	*02:01	*34:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
24	9035 JBUSH	*32:01			-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
25	9049 IBW9	*33:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
26	9285 WT49	*02:05			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
27	9191 CH1007	*24:10	*29:01		+	-	-	-	-	-	-	-	-	-	-	-	+	+	-	-
28	9320 BEL5GB	*02:01	*29:02		-	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-
29	9050 MOU	*29:02			-	-	+	-	-	-	-	-	-	-	-	-	+	+	-	-
30	9021 RSH	*30:01	*68:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
31	9019 DUCAF	*30:02			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
32	9297 HAG	*02:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
33	9098 MT14B	*31:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
34	9104 DHIF	*31:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
35	9302 SSTO	*32:01			-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
36	9024 KT17	*02:06	*11:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
37	9065 HHKB	*03:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
38	9099 LZL	*02:17			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
39	9315 CML	*01:01	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
40	9134 WHONP199	*02:07	*30:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
41	9055 H0301	*03:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
42	9066 TAB089	*02:07			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076 T7526	*02:06	*02:07		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057 TEM	*66:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
45	9239 SHJO	*23:01	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
46	9013 SCHU	*03:01			-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045 TUBO	*02:16	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
48	9303 TER-ND	*02:01	*11:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

101.428-12 – including Taq polymerase, IFU-01
101.428-12u – without Taq polymerase, IFU-02

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“Instructions for Use” (IFU)

Lot No.: 5L0

Lot-specific Information

CELL LINE VALIDATION SHEET																	
HLA-A*29 SSP subtyping kit ²																	
				Well													
				17	18	19	20	21	22	23	24	25	26	27	28	29	30
				201901617	201901618	201901619	201901620	201901621	201901622	201901623	201901624	201901625	201901626	201901627	201901628	201901629	202023730
	IHWC cell line ¹	A*	A*	Lot No.:													
1	9001 SA	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280 LK707	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	9011 E4181324	*01:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275 GU373	*30:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
5	9009 KAS011	*01:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353 SM	*02:01	*26:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020 QBL	*26:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
8	9007 DEM	*31:01		-	-	-	-	-	-	+	-	-	-	-	-	-	-
9	9026 YAR	*26:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107 LKT3	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
11	9051 PITOUT	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
12	9052 DBB	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004 JESTHOM	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071 OLGA	*31:01		-	-	-	-	-	-	+	-	-	-	-	-	-	-
15	9075 DKB	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037 SWEIG007	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
17	9282 CTM3953540	*03:01	*80:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
18	9257 32367	*33:03	*74:01	-	-	-	+	-	-	+	-	-	-	-	-	-	-
19	9038 BM16	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
20	9059 SLE005	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
21	9064 AMALA	*02:17		-	-	-	-	-	-	-	-	-	-	-	-	-	-
22	9056 KOSE	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
23	9124 IHL	*02:01	*34:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
24	9035 JBUSH	*32:01		-	-	-	+	-	-	+	-	-	-	-	-	-	-
25	9049 IBW9	*33:01		-	-	-	-	-	-	+	-	-	-	-	-	-	-
26	9285 WT49	*02:05		-	-	-	-	-	-	-	-	-	-	-	-	-	-
27	9191 CH1007	*24:10	*29:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
28	9320 BEL5GB	*02:01	*29:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-
29	9050 MOU	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
30	9021 RSH	*30:01	*68:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-
31	9019 DUCAF	*30:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
32	9297 HAG	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
33	9098 MT14B	*31:01		-	-	-	-	-	-	+	-	-	-	-	-	-	-
34	9104 DHIF	*31:01		-	-	-	-	-	-	+	-	-	-	-	-	-	-
35	9302 SSTO	*32:01		-	-	-	+	-	-	+	-	-	-	-	-	-	-
36	9024 KT17	*02:06	*11:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
37	9065 HHKB	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
38	9099 LZL	*02:17		-	-	-	-	-	-	-	-	-	-	-	-	-	-
39	9315 CML	*01:01	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
40	9134 WHONP199	*02:07	*30:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
41	9055 H0301	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
42	9066 TAB089	*02:07		-	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076 T7526	*02:06	*02:07	-	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057 TEM	*66:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
45	9239 SHJO	*23:01	*24:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-
46	9013 SCHU	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045 TUBO	*02:16	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
48	9303 TER-ND	*02:01	*11:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-

101.428-12 – including *Taq* polymerase, IFU-01
 101.428-12u – without *Taq* polymerase, IFU-02

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Lot No.: **5L0**

Lot-specific Information

¹The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 2, 4 to 7, 9 to 11, 13, 17 to 19, 21, 22 and 24 to 30 were available.

The specificities of the primers in primer solutions 4, 6, 10, 11, 13, 17, 18, 21, 22 and 29 were tested by separately adding one, two or three additional 5'-primers, and one or two additional 3'-primers respectively.

In primer solutions 2, 5, 7, 9, 19, 24 to 26 and 30 it was only possible to test the 5'-primers, the 3'-primers were not possible to be tested.

In primer solutions 27 and 28 it was only possible to test the 3'-primer, the 5'-primer was not possible to be tested.

In primer solutions 4, 6, 8, 11 to 13, 17, 18, 22 and 29 one or more of the 3'-primers were not possible to be tested, and in primer solutions 10, 16, 17 and 23 one, two or three of the 5'-primers were not possible to be tested. In addition, one or more primers in primer solutions 8 and 16 were tested by separately adding one 3'-primer and one 5'-primer respectively.

101.428-12 – including *Taq* polymerase, IFU-01
 101.428-12u – without *Taq* polymerase, IFU-02

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 “Instructions for Use” (IFU)

Lot No.: **5L0**

Lot-specific Information

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